AD	

Award Number: DAMD17-00-1-0238

TITLE: Development of Dual Acting Inhibitors for Breast Cancer

PRINCIPAL INVESTIGATOR: Pui-Kai Li, Ph.D.

CONTRACTING ORGANIZATION: The Ohio State University

Columbus, Ohio 43210-1239

REPORT DATE: August 2002

TYPE OF REPORT: Annual

PREPARED FOR: U.S. Army Medical Research and Materiel Command

Fort Detrick, Maryland 21702-5012

DISTRIBUTION STATEMENT: Approved for Public Release;

Distribution Unlimited

The views, opinions and/or findings contained in this report are those of the author(s) and should not be construed as an official Department of the Army position, policy or decision unless so designated by other documentation.

	REPORT DOCUMENTATION PAGE			Form Approved DMB No. 074-0188
	mation is estimated to average 1 hour per respons this collection of information. Send comments reg rs Services, Directorate for Information Operations Project (0704-0188) Washington DC 2050		structions, searching	existing data sources, gathering and maintaining
1. AGENCY USE ONLY (Leave blan	nk) 2. REPORT DATE	3. REPORT TYPE AND		
4. TITLE AND SUBTITLE	August 2002	Annual (3 Jul		
	1 Astine Tehilitary		5. FUNDING	
	l Acting Inhibitors	s for Breast	DAMD17-00	0-1-0238
Cancer				
6. AUTHOR(S)				
Pui-Kai Li, Ph.D.				
·				
7. PERFORMING ORGANIZATION I	NAME(S) AND ADDRESS(ES)		8. PERFORMII	NG ORGANIZATION
m1 01			REPORT NUMBER	
The Ohio State Uni				
Columbus, Ohio 43	210-1239			
E-Mail: li.27@osu.edu				
9. SPONSORING / MONITORING A	AGENCY NAME(S) AND ADDRESS(E	S)	10 SPONSOR	INC / MONITODING
		,	10. SPONSORING / MONITORING AGENCY REPORT NUMBER	
U.S. Army Medical Research and	d Materiel Command			
Fort Detrick, Maryland 21702-5	012			
44 CUDDI FAIGHTA DV NOTES				
11. SUPPLEMENTARY NOTES				
12a. DISTRIBUTION / AVAILABILIT	Y STATEMENT			12b. DISTRIBUTION CODE
	elease; Distribution Un	limited		12b. DISTRIBUTION CODE
				-
13. ABSTRACT (Maximum 200 W	ords)			1
Purpose: To design dual acting inh	nibitors that can block the enzyme e	strone sulfatase (an enzyn	ne involves in t	he in situ formation of
estrogen in breast cancer cells) and	act as antiestrogens.			
Scope: The design and synthesis of	f 30 dual inhibitors are proposed. Th	he inhibitors contain 4 diff	ferent structural	core. The synthesized
initiotions will be tested on their ab	lifty to inhibit the enzyme estrone si	ultatase and also their ahil	ity to inhihit th	e growth of breact
cancer cells stimulated by estrone s	sulfate. In addition, selected inhibito	ors will be tested in vivo us	sing NMU-indu	iced mammary tumors
in rats.				-
been tested for their ability to inhib	16 out of 30) of the proposed inhibitit estrone sulfatase, activity of rat li	tors have been synthesized	d. Six out of the	e 16 inhibitors have
20 min of substrate estrone sufface.	All the inhibitors tested so far are m	nore notent than our lead o	rompound Tom	ovifon gulformete
Naioxiiche sunamate (minottor 30)	exhibits an extremely notent sulfat	ase inhibitory activity. It is	nhihite more th	on 050/ of the gulfatage
activity at 20 pivi concentration, it	is by far the most potent qual innibi	tor we have ever obtained	. It may serves	as an important new
icad in scarch of more potent and e	ffective dual inhibitors for the treatr	ment of breast cancer.		
44 CUD FOT TERMS				
14. SUBJECT TERMS breast cancer, dual inhibitors			15. NUMBER OF PAGES	
breast cancer, dual inhibitors		<u> </u>	15	
				16. PRICE CODE
17. SECURITY CLASSIFICATION	18. SECURITY CLASSIFICATION	19. SECURITY CLASSIFI	CATION	20. LIMITATION OF ABSTRACT
OF REPORT	OF THIS PAGE	OF ABSTRACT		
Unclassified	Unclassified	Unclassifi	ed	Unlimited
NSN 7540-01-280-5500			Stan	dard Form 208 (Pov. 2-90)

Table of Contents

Cover 1
SF 298 2
Table of Contents 3
Introduction4
Body 5-7
Key Research Accomplishments 8
Reportable Outcomes9
Conclusions10
References11
Appendices12-1

Introduction

Breast cancer is the most common malignancy in the United States. It is estimated that approximately 30 -40 % of all breast cancers are estrogen-dependent. Currently, the most common treatments use either antiestrogen or aromatase inhibitors. They are effective in 35-40 % of advanced postmenopausal breast cancer patients. In estrogen-dependent breast cancer patients, the estrogen levels in breast cancer cells are 5-10 times higher than in plasma. One of the possibilities to explain this observation is in situ production of estrogens from precursor substrates in the breast cancer cells. One of the pathways for the in situ production of estrogen is the conversion of androgens to estrogens by the enzyme aromatase (aromatase pathway). Another pathway for the in situ formation of estrogen is through the conversion of estrone sulfate to estrone by the enzyme estrone sulfatase (estrone sulfatase pathway). It has been pointed out that the estrone sulfatase pathway is significant and produce 10 times more estrogen than through the aromatase pathway in breast cancer cells. In addition, estrone sulfatase is also responsible for the conversion of dehydroepiandrosterone sulfate to androst-5-ene-3\(\beta\),17\(\beta\)-diol, another estrogenic steroid in the body. Thus, potent estrone sulfatase inhibitors are potential agents for the treatment of estrogendependent breast cancer. Preliminary studies demonstrated that estrone sulfatase inhibitor can block the growth of NMU-induced tumor in rat stimulated by estrone sulfate. Thus the current approach is to design dual acting inhibitors that can not only block the estrone sulfatase pathway, but also act as antiestrogens. The proposed dual acting inhibitors will have advantage over the current drug treatments. The inhibitors will not only block the formation of estrogen, but also block the stimulatory effect of estrogen on cancer cells. This proposal will design and synthesize of dual acting inhibitors with sulfatase inhibitory and anti-estrogenic activity. The synthesized inhibitors will be tested using enzyme inhibition and cell culture assays. Finally, In vivo studies of dual acting inhibitors using NMU-induced mammary tumor in rats will be performed.

Body

As stated in the introduction, this proposal deals with the design, synthesis and biological testings of dual inhibitors with sulfatase inhibitory and anti-estrogenic activities. A total of 30 inhibitors are proposed. In the first year we have synthesized 16 inhibitors (inhibitors 1-15 and 30 stated in the proposal). This year, we have completed the synthesis of inhibitors 16-24 proposed in the grant. We have completed 83 % of the proposed compounds. The synthesis of the remaining compounds (inhibitors 25-29) should be completed in the near future.

The syntheses of inhibitors 16-24 are shown below (Fig. 1).

The syntheses of inhibitors 16-24 are shown in Fig. 1. Inhibitors 16-24 are analogs of inhibitors 7 – 15 with a benzocycloheptene nucleus instead of a dihydronaphthalene. Compound 1 is one of the starting material which was prepared from m-anisaldehyde in 3 steps by the published procedure (ref 1). The methyl group in 1 was replaced by a tetrahydropyranyl group to form compound 2. Treatment of 3 with n-butyllithium, then with ketone 2), followed by dehydration of the resulting tertiary alcohol with silica gel, afforded olefin 4 (57.5 % based on 4). Bromination of compound 4 with pyridinium tribromide followed by acidic hydrolysis furnished the vinyl bromide 5 (92.6 %). Palladium catalyzed coupling of compound 5 with various para-substituted phenyl zinc chlorides which were prepared by the treatment of the corresponding substituted phenylbromides with n-butyl lithium followed by zinc chloride, gave compounds 30-38 (84-90%). Iodination of alcohols 30-38 with L/PPh₂/Imidazole yielded the iodides 39-47 (90-94%). Reaction of compound 39-47 with dimethylamine gave the corresponding amines 48-56 respectively (80-85 -91%). Sulfamoylation (ref 2) of 48-49, 51-56 with sulfamovl chloride in the presence of hinder base: 2,6-di-tert-butyl-4-methyl pyridine, yielded the target compounds 16-17, 19-24, respectively (81-87%). Hydrogenation of compound **50** yielded inhibitor **18** (87%).

The syntheses of inhibitors 16-24 are similar to the synthesis of inhibitors 7-15 and we did not come across any difficult.

Figure 1. Synthesis of Inhibitors 16-24

Reagents and Conditions a. i) BBr₃, CH₂Cl₂, r.t, 4 h; ii) Dihydropyran, PPTs, CH₂Cl₂, r.t, 2.5 h, 98 %; b. i) n-BuLi, THF, -78°C, 45 min; ii) 4, -78°C to r.t, 3 h; iii) SiO₂, CH₂Cl₂, r.t, overnight, 57.5 % based on 4; c. i) C₅H₅N.HBr₃, CH₂Cl₂, 0°C, 1.5 h; ii) 2N HCl, THF, r.t, 1.5 h, 92.6 %; d. R-Ph-ZnCl (R = OCH₃, CH₃, OCH₂Ph, C₂H₅, n-C₃H₇, i-C₃H₇, n-C₄H₉, i-C₄H₉, t-C₄H₉), Pd(PPh₃)₄, THF, reflux, 2.5 h, 84 - 90 %; e. I₂, PPh₃, Imidazole, CH₂Cl₂, r.t, 40 min, 90 - 94 %; f. (CH₃)₂NH, K₂CO₃, THF, r.t, 20 h, 80-85%; g. ClSO₂NH₂, 2,6-di-tert-butyl-4-methylpyridine, r.t, 1 h, 81-87%; h. H₂, 10% Pd/C, CH₂Cl₂-CH₃OH (3:1), r.t, 1 h, 87 %.

Enzyme Inhibition studies of inhibitors

Inhibitors 16-24 were tested for their abilities to inhibit estrone sulfatase activity of rat liver microsomes at 20 μ M concentrations and in the presence of 20 μ M substrate estrone sulfate. The sulfatase inhibitory activities of the inhibitors are similar to inhibitors 7-15. At 20 μ M inhibitor concentration, the % inhibiton of sulfatase activity activity range from 35-55%. Raloxifene sulfamate (inhibitor 30) is still the most potent inhibitor among all the inhibitors we synthesized (over 95% inhibition) at the same inhibitor concentration. We have synthesized 4 grams of the inhibitors for in vivo anti-tumor study.

Key Research Accomplishment

More than 83% (26 out of 30) of the proposed inhibitors have been synthesized All the inhibitors tested so far are more potent than our lead compound Tamoxifen sulfamate. Raloxifene sulfamate (inhibitor 30) exhibits an extremely potent sulfatase inhibitory activity. It inhibits more than 95% of the sulfatase activity at 20 μ M concentration. It is by far the most potent dual inhibitor we have ever obtained. We have synthesized 4 grams of the inhibitors for in vivo antitumor study.

Reportable Outcomes

- 1. A manuscript has been prepared on the synthesis and sulfatase inhibitory activities of dual inhibitors with nafoxidine nucleus (please refer to the appendix)
- 1. Currently a research associate is involved in the synthesis of and a second year graduate student on the biological testing of the inhibitors.

Conclusions:

Twenty-five out of the 30 proposed inhibitors have been synthesized and tested for their ability to inhibit estrone sulfatase activity of rat liver microsomes at 20 μ M concentrations and in the presence of 20 μ M substrate estrone sulfate. The inhibitors belong to the nafoxidine, benzocyclohepterne and raloxifene structural classes. All the inhibitors showed significant inhibition of estrone sulfatase and are more potent than our lead compound Tamoxifen sulfamate. Raloxifene sulfamate (inhibitor 30) exhibits an extremely potent sulfatase inhibitory activity and has been chosen to be one of the compounds for in vivo anti-tumor study. We have synthesized 4 grams of the compound.

Reference

1. R. McCague, G. Leclercq and C. Jordan. Nonisomerizable analogues of (Z)- and (E)-4-hydroxytamoxifen. Synthesis and endocrinological properties of substituted diphenylbenzocycloheptenes. J. Med. Chem. 31: 1285-1290 (1988).

SYNTHESIS AND SULFATASE INHIBITORY ACTIVITIES OF CONFORMATIONAL RESTRICTED ANALOGS OF (E) -4-HYDROXYTAMOXIFEN SULFAMATE

Guo-Hua Chu⁺, Heidi Kabler, Kyle W. Selcer[#], Jiandong Shi, Zhigen Hu⁺ and Pui-Kai Li^{+*}

[†]Division of Medicinal Chemistry and Pharmacognosy, College of Pharmacy, The Ohio State University, Columbus OH, 43210

> *Department of Biology, Bayer School of Natural and Environmental Sciences, Duquesne University, Pittsburgh PA 15282

Abstract: Eight conformational restricted analogs of (E)-4-hydroxytamoxifen sulfamate are synthesized as estrone sulfatase inhibitors. All the inhibitors significantly inhibited estrone sulfatase activity. Varying the nature of the substituents in R_3 (H, CH_3 , OCH_3 , OH) has little effect on the sulfatase inhibitory activity. However, inhibitors with pyrrolidinyl group consistently exhibit higher sulfatase inhibitory activities than the inhibitors with dimethylamino groups.

There is substantial evidence that breast tumors in post-menopausal women accumulate high concentration of estrogens^{1,2} and possibly through conversion of estrone sulfate to estrone by the enzyme estrone sulfatase.^{3,4} Several estrone sulfatase inhibitors (both steroidal and non-steroidal) have been developed as potential agents for the treatment of estrogen-dependent breast cancers.^{5,26} Since the pharmacophore for sulfatase inactivation is a phenylsulfamoyl group, it occurs to us that a potent antiestrogen such as (Z) 4-hydroxytamoxifen can be easily converted to the respective sulfamate analog and becomes potential dual inhibitor (inhibitor with sulfatase inhibitor activity and antiestrogenic activity). Recently, we have synthesized (E) 4-hydroxytamoxifen sulfamate (Fig. 1), the sulfamoylated analog of the potent antiestrogen (Z) 4-hydroxytamoxifen, and demonstrated that it is four fold better than the substrate estrone sulfate in binding to estrone sulfatase.²⁷ However, the potent antiestrogen (Z)-4-hydroxytamoxifen has been shown in vitro to isomerize into a mixture of Z and E isomer.²⁸ (E)-4-Hydroxytamoxifen with the central double bond is responsible for the facile isomerization [103].³⁰, one method to fix the configuration of the double bond is incorporating it into a ring such as in nafoxidine. Thus, inhibitors 1 - 8 were synthesized as conformational restricted analogs of (E) 4-hydroxytamoxifen sulfamate.

(E)-4-Hydroxytamoxifen sulfamate

1, $R_1=R_2=CH_3$, $R_3=H$ 3, $R_1=R_2=CH_3$, $R_3=OCH_3$ 2, $R_1=R_2=R_3=CH_3$, 32, $R_1=R_2=CH_3$, $R_3=OCH_2Ph$

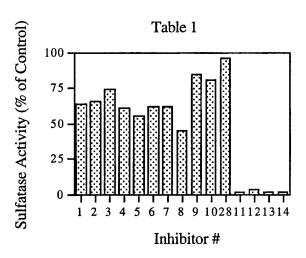
 $5, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = H \\ 7, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = OCH_{3} \\ 6, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = CH_{3} \\ 33, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = OCH_{2}Ph \\ 29, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = CH_{3} \\ 31, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = OCH_{2}Ph \\ 29, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = CH_{3} \\ 31, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = OCH_{2}Ph \\ 29, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = CH_{3} \\ 31, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = OCH_{2}Ph \\ 29, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = CH_{3} \\ 31, R_{1}, R_{2} = -(CH_{2})_{4}, R_{3} = CH_{3} \\$

$$H_2NO_2SO$$
 $32,33$
 H_2NO_2SO
 $A,8$

Reagents and Conditions: a. BrCH₂CO₂Et, K₂CO₃, acetone, reflux 2.5 h, 99.3 %; b. LiAlH₄, THF, r.t, 2 h; c. TBSCl, Imidazole, DMF, r.t, overnight, 96.4 % for 2 steps; d. Dihydropyran, PPTs, CH₂Cl₂, r.t, 2.5 h, 98 %; e. i) n-BuLi, THF, -78°C, 45 min; ii) 13, -78°C to r.t, 3 h; iii) SiO₂, CH₂Cl₂, r.t, overnight, 65.7% based on 13; f. i) C₅H₅N.HBr₃, CH₂Cl₂, 0°C, 1.5 h; ii) 2N HCl, THF, r.t, 1.5 h, 90.3 %; g. R-Ph-ZnCl (R = H, CH₃, OCH₃, OCH₂Ph), Pd(PPh₃)₄, THF, reflux, 2.5 h, 91 - 94 %; h. I₂, PPh₃, Imidazole, CH₂Cl₂, r.t, 40 min, 93 - 95 %; i. (CH₃)₂NH or pyrrolidine, K₂CO₃, THF, r.t, 20 h, 88.3 - 94.1%; j. ClSO₂NH₂, 2,6-di-tert-butyl-4-methylpyridine, r.t, 1 h, 91 - 94 %; k. H₂, 10% Pd/C, CH₂Cl₂-CH₃OH (3:1), r.t, 1 h, 79.2 % for 4, 82 % for 8.

The synthesis of compounds 1 - 8 is summarized in scheme 1. Reaction of 4-bromophenol 9 with ethyl bromoacetate gave ester 10 (99.3 %). Reduction of 10 with LiAlH₄ followed by protection of the resulting alcohol 11 as TBS ether yielded compound 12 (96.4 % for 2 steps). Treatment of 12 with n-butyl lithium, then with ketone 13 which was prepared by tetrahydropyranylayion of 6-hydroxy-1-tetralone (98%), followed by dehydration of the resulting tertiary alcohol with silica gel, afforded olefin 14 (65.7% based on 13). Bromination of compound 14 with pyridinium tribromide followed by acidic hydrolysis furnished the vinyl bromide 15 (90.3%). Palladium catalyzed coupling³¹ of compound 15 with various para-substituted phenyl zinc chlorides which were prepared by the treatment of the corresponding substituted phenylbromides with n-butyl lithium followed by zinc chloride, gave compounds 16 - 19 (91-94%). Iodination of alcohols 16 - 19 with I₂/PPh₃/Imidazole yielded the iodides 20 - 23 (93-95%). Reaction of compounds 20 - 23 with dimethylamine or pyrrolidine gave the corresponding amines 24 - 27 and 28 - 31 respectively (88.3 -94.1%). Sulfamoylation³² of 24 -26 and 28 - 30 with sulfamoyl chloride in the presence of hinder base: 2,6-di-tert-butyl-4-methyl pyridine, yielded the target compounds 1 - 3 and 5 - 7 respectively. Compound 4 was synthesized by sulfamoylation of compound 27 to form compound 32 followed by debenzylation through hydrogenation. The synthesis of inhibitor 8 was similar to 4 except compound 31 was sulfamoylated instead.

Inhibitors 1 - 8 can be divided into 2 series. Both series have the same modifications at the para position (R_3) of the 2-phenyl group. Series 1 (inhibitors 1 - 4) contain the dimethylamino ethyl moiety while series 2 (inhibitor 5 - 8) have the pyrrolidinyl ethyl moiety. Inhibitors 1 - 8 were tested for their ability to inhibit estrone sulfatase activity of rat liver microsomes at 20 μ M concentrations and in the presence of 20 μ M substrate estrone sulfate. Table 1 shows the relative inhibition of estrone sulfatase by the inhibitors. All the inhibitors significantly inhibited estrone sulfatase activity. The sulfamate moiety is essential for sulfatase inhibition since compound 24, the precursor of inhibitor 1, did not show sulfatase inhibitory activity (Table 1). Varying the nature of the substituents in R_3 (H, CH_3 , OCH_3 , OH) has little effect on the sulfatase inhibitory activity. However, inhibitors with pyrrolidinyl group (inhibitors 5 - 8) consistently exhibit higher sulfatase inhibitory activities than the inhibitors with dimethylamino groups (inhibitor 1 - 4).



References and Notes

- 1. Noel, C.T.; Reed, M.J.; Jacobs, H.S.; James, V.H.T. J. Steroid Biochem. 1981, 14, 1101.
- 2. Samoilik, E.; Santen, R.J.; Worgul, T.J. Steroids 1982, 39, 497.
- 3. Santner, S.J.; Feil, P.D.; Santen, R.J. J. Clin. Endocrinol. Metab. 1984, 59, 29.
- 4. Pasqualini, J.R.; Chetrite, G.; Nguyen, B.L.; Maloche, C.; Delalonde, L.; Talbi, M.; Feinstein, M.C.; Blacker, C.; Botella. J.; Paris, J. J. Steroid Biochem. & Molec. Biol. 1995, 59, 407.
- 5. Duncan, L.; Puroit, A.; Howarth, N.M.; Potter, B.V.L.; Reed, M.J. Cancer Res. 1993, 53, 298.
- 6. Howarth, N.M.; Cooper, G.; Purohit, A.; Duncan, L.; Reed, M.J.; Potter, B.V.L. Bioorg. Med. Chem. Lett. 1993, 3, 313.
- 7. Li, P.K.; Pillai, R.; Young, B.L.; Bender, W.H.; Martino, D.M.; Lin, F.T. Steroids 1993, 58, 106.
- 8. Howarth, N.M.: Purohit, A.; Reed, M.J.; Potter, B.V.L. J. Med. Chem. 1994, 37, 219.
- 9.Dibbelt, L.; Li, P.K.; Pillai, R.; Knuppen, R. J. Steroid Biochem. Molec. Biol. 1994, 50, 261.
- 10. Dibbelt, L.; Li, P.K.; Pillai, R.; Knuppen, R. J Steroid Biochem. & Molec. Biol. 1995, 52, 281.
- 11. Purohit, A.; Williams, G.J.; Howarth, N.M.; Potter, B.V.L.; Reed, M.J. Biochemistry 1995, 34, 11508.
- 12. Purohit, A.; Williams, G.J.; Roberts, C.J.; Potter, B.V.L.; Reed, M.J. Int. J. Cancer 1995, 63, 106.
- 13. Li, P.K.; Pillai, R.; Dibbelt, L. Steroids 1995, 60, 299.
- 14. Li, P.K.; Milano, S.; Kluth, L.; Rhodes, M.E. J. Steroid Biochem. & Molec. Biol. 1996, 59, 41.
- 15. Selcer, K.W.; Jagannathan, S.; Rhodes, M.E.; Li, P.K. J. Steroid Biochem. & Molec. Biol. 1996, 59, 83.
- 16. Woo, L.W.; Lightowler, M.; Purohit, A.; Reed, M.J.; Potter, B.V.L. J Steroid Biochem. & Mol. Biol. 1996, 57, 79.
- 17. Woo, L.W.; Purohit, A.; Reed, M.J.; Potter, B.V.L. J Med. Chem. 1996, 39, 1349.
- 18. Selcer, K.W.; Hedge P.; Li, P.K. Cancer Res. 1997, 57, 702.
- 19. Chu, G.H.; Milano, S.; L. Kluth, Rhodes, M.E.; Johnson, D.A.; Li, P.K. Steroids 1997, 62, 530.
- 20. Li. P.K.: Chu, G.H.; Peters, A.; Selcer, K.W. Steroids 1998, 63, 425.
- 21. Poirier, D.; Boivin, R.P. Bioorg. Med. Chem. Lett. 1998, 8, 1891.
- 22. Purohit, A.; Vernon, K.A.; Hummelinck, A.E.; Woo, L.W.; Hejaz, H.A.; Potter, B.V.; Reed, M.J. J Steroid Biochem. Mol. Biol. 1998, 64, 269.
- 23. Woo, L.W.; Howarth, N.M.; Purohit, A.; Hejaz, H.A.; Reed, M.J.; Potter, B.V.L. *J Med. Chem.* 1998, 41, 1068.
- 24. Kolli, A.; Chu, G.H.; Rhodes, M.E.; Inoue, K.; Selcer, K.W.; Li, P.K. *J Steroid Biochem. Mol. Biol.* **1999**, 68, 31.
- 25. Ciobanu, L.C.; Boivin, R.P.; Luu-The, V.; Labrie, F.; Poirier, D. J Med. Chem. 1999, 42, 2280.
- 26. Hejaz, H.A.; Purohit, A.; Mahon, M.F.; Reed, M.J.; Potter, B.V. J Med. Chem. 1999, 42, 3188.
- 27. Chu, G.H.; Peters, A.; Selcer, K.W.; Li, P.K. Bioorg. Med. Chem. Lett. 1999, 9, 141.
- 28. Katzenellenbogen, J.A.; Carlson, K.E.; Katzenellenbogen, B.S. J. Steroid Biochem., 1985, 22, 589.
- 29. Harper, M.J.; Walpole, A.L. Nature 1966, 212, 87.
- 30. Jordon, V.C.; Haldemann, B.; Allen, K.E. Endocrinology 1981, 108, 1353.
- 31. McCague, R.; Leclercq, G.; Jordan, V.C.; J med. Chem. 1988, 31, 1285.
- 32. Schwartz, S.; Thieme, I.; Richter, M.; Undeutsch, B.; Henkel, H.; Elger, W. Steroids, 1996, 61, 710.